

Review Article

Update on the Cardiovascular Risk in Obesity: Endocrine and Paracrine Role of the Adipose Tissue

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The prevalence and incidence of obesity, an important cardiovascular risk factor, are increasing dramatically in Greece and worldwide. This article highlights recent insights into the role of adipose tissue-derived factors in mediating the vascular complications of obesity, particularly atherosclerosis.

The prevalence of obesity in Greece and worldwide

The mean body weight and prevalence of obesity in the population have increased alarmingly over the past two to three decades. According to the World Health Organization (WHO), 200 million adults worldwide were obese (defined as body mass index [BMI] >30 kg/m²) in 1995, whereas in 2005 approximately 1.6 billion people were overweight (BMI 25-29.9 kg/m²) and at least 400 million obese (www.who.int/mediacentre/factsheets/fs311/en). Projected estimates for 2015 indicate that 2.3 billion individuals will be overweight and more than 700 million obese.

A recently conducted cross-sectional nationwide survey in Greece included 17,341 men and women aged from 20 to 70 years and indicated (based on self-reported data) that the overall prevalence of overweight and obesity amounts to 35.2%

and 22.5%, respectively.¹ Abdominal obesity (defined as waist circumference >102 cm in males and >88 cm in females) was present in 41.1% of men and 29.9% of women. Even higher percentages were reported in earlier, smaller studies. For example, the cross-sectional analysis of 4153 adult participants (older than 18 years) of the MetS-Greece Multicentre Study (data collected during 2003) revealed a high prevalence (56.8%) of abdominal obesity,² while the European Prospective Investigation into Cancer and Nutrition (EPIC) study (data collected between 1992-2000) found that abdominal obesity in Greece was present in 38.8% of men and 54.5% of women aged 50 to 64 years.³ Furthermore, evaluation of more than 3000 persons (age 20-89 years) living in the Attica region between 2001 and 2002 revealed that 53% of men were overweight and 20% obese; for women, the rates were 31% and 15%, respectively.⁴

Overall, these numbers demonstrate the magnitude of the health problem associated with obesity. They also show that deeper insights into the molecular pathways and cardiovascular consequences of increased body weight are urgently needed to prevent and combat the impact of increased body fat mass on cardiovascular morbidity and mortality in the near future.

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Obesity as cardiovascular risk factor

Obesity is a major risk factor for coronary heart disease⁵ and clinical studies have shown that it increases the risk of suffering or dying from atherothrombotic complications such as myocardial infarction or stroke.⁶ Obese persons have a higher incidence of arterial hypertension and left ventricular hypertrophy, both important risk factors for the development of heart failure.⁷ Based on these facts, statistical forecasts have estimated that obesity-associated cardiovascular mortality will soon neutralize the increase in life expectancy achieved over the past decades as a consequence of improvements in living conditions and medical care.⁸

Important insights into potential pathomechanisms underlying the increased cardiovascular risk in obesity have been obtained over the past years; however, the molecular pathways are far from being completely understood. An increase in visceral adipose tissue mass is frequently accompanied by metabolic alterations such as hyperinsulinemia, glucose intolerance and diabetes, as well as hyper- or dyslipidemia, all of which may promote the development of atherosclerosis. In addition, factors produced and released from the adipose tissue may directly modulate the function of vascular cells and contribute to the formation of atherosclerotic lesions in obesity. The effects of some of these so-called adipokines on vascular cells and vascular lesion formation are discussed in more detail below.

Adipose tissue as source of inflammatory cytokines

Obesity is accompanied by an increased expression of adhesion receptors on adipocytes, followed by an enhanced infiltration of the adipose tissue with inflammatory cells, primarily macrophages.⁹ Adipose tissue macrophages, which may constitute up to 40% of all cells within the adipose tissue, are an important source of proinflammatory cytokines, such as tumor necrosis factor (TNF) α , interleukin (IL)1, IL6 or monocyte chemoattractant protein 1/CC-chemokine ligand 2 (MCP1/CCL2) (Figure 1); these not only contribute to the systemic proinflammatory condition frequently associated with obesity, but may also act locally and adversely affect adipocyte function, e.g. promote the development of insulin resistance. For instance, experimental studies in obese mice have shown that lack of the receptor for MCP1/CCL2 not only reduced adipose tissue inflammation, but also improved insulin resistance.¹⁰ Furthermore, macrophage-secreted chemokines (e.g. MCP1/CCL2 and IL8/CXC ligand 8 [CXCL8]) as well as adipokines (e.g. leptin) are potent angiogenic growth factors, and an enhanced vascularization of the adipose tissue could further promote obesity and its metabolic complications by facilitating inflammatory cell recruitment. In agreement with these data, inhibition of angiogenesis was shown to reduce adipose tissue expansion and the development of obesity.¹¹ Reciprocally, binding of products released from adipocytes (e.g. free fatty

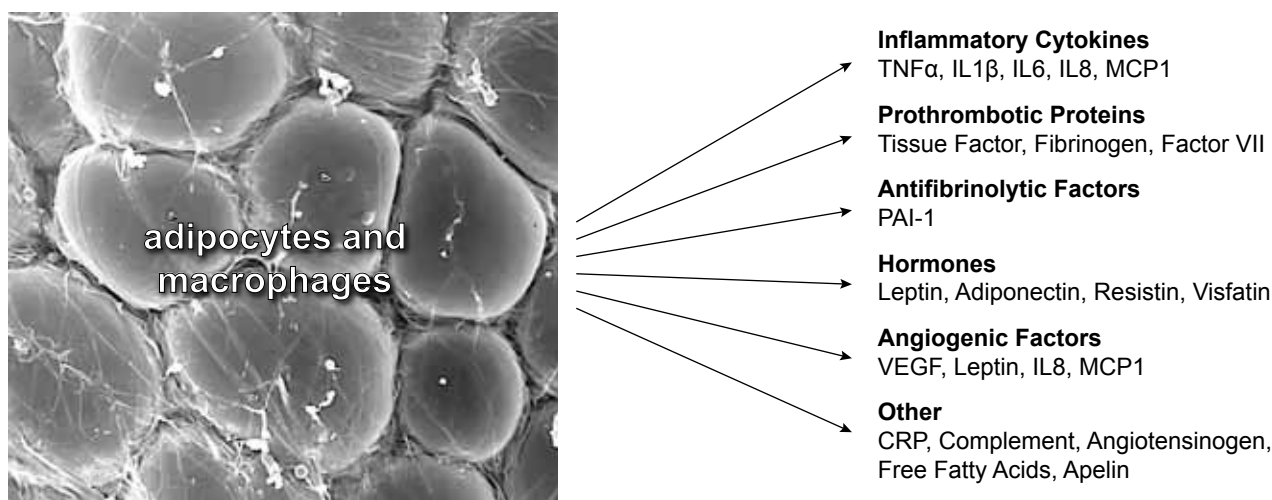


Figure 1. Obesity is associated with local and systemic inflammation. List of factors produced in and secreted from adipose tissue (adipocytes or adipose tissue-resident macrophages), grouped according to their major function. CRP – C-reactive protein; IL – interleukin; MCP-1 – monocyte chemoattractant protein-1; PAI-1 – plasminogen activator inhibitor-1; TNF α – tumor necrosis factor-alpha; VEGF – vascular endothelial growth factor.

acids) to receptors expressed on macrophages (e.g. Toll-like receptors) may lead to the subsequent activation of the latter cell type, ultimately resulting in a self-perpetuating vicious circle of macrophage and adipocyte stimulation.

The chronic, low-grade inflammation associated with obesity is reflected by elevated levels of the acute phase reactant C-reactive protein (CRP),¹² produced in the liver as a result of chronic stimulation with IL6 released from the adipose tissue. In addition, there is evidence that CRP is produced in the adipose tissue itself,¹³ and plasma CRP levels correlate with body fat mass and indices of visceral adiposity,¹⁴ whereas surgically or diet-induced weight loss are associated with reduced CRP levels.¹⁵ Interestingly, serum CRP is not only a potential biomarker of increased cardiovascular risk,¹⁶ but also appears to possess direct effects within the cardiovascular system. For example, experimental evidence suggests that CRP promotes angiogenesis¹⁷ or thrombosis,¹⁸ and CRP transgenic apolipoprotein E knockout mice exhibited accelerated atherosclerosis lesion formation.¹⁹ However, opposite findings have also been reported.^{20,21} Thus, the role of CRP as an active player within the cardiovascular system remains undetermined.

Cardiovascular risk associated with different adipose tissue depots

Clinical studies have found visceral adiposity, rather than the enlargement of subcutaneous fat depots, to be associated with an increased cardiovascular risk. Possible mechanisms include the direct drainage of adipokines and other metabolites from intra-abdominal fat depots into the portal circulation (followed by stimulation of the hepatic production of CRP, plasminogen activator inhibitor-1, or fibrinogen),²² but also the higher expression of proinflammatory mediators²³ and stronger infiltration with macrophages⁹ compared to subcutaneous adipose tissue. In addition, recent studies point to a role for other fat depots, e.g. those surrounding blood vessels or within the pericardium, in the pathogenesis of atherosclerosis in obesity (Figure 2). For example, the amount of intrathoracic or pericardial fat was found to correlate with the severity of coronary artery calcification,²⁴ or the incidence of coronary heart disease and myocardial infarction.²⁵ Interestingly, perivascular fat volumes >300 cm³ were associated with a higher relative risk for coronary atherosclerosis than traditional systemic cardiovascular risk

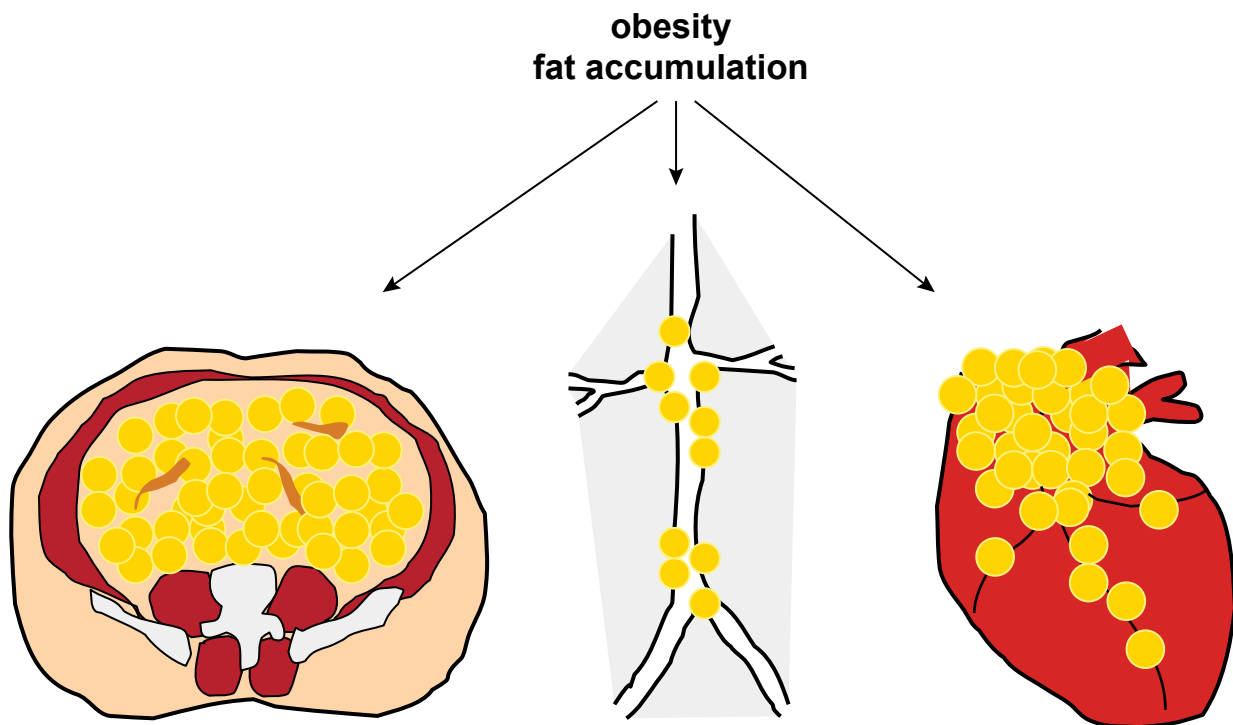


Figure 2. Fat deposition in obesity. Obesity is associated with increased fat deposition within the abdominal cavity, but also around blood vessels or within the pericardium.

Table 1. Short summary of some major effects of adipokines potentially relevant to vascular lesion formation in obesity.

	Findings in mouse models or vascular cells	References
Adiponectin	– increases insulin sensitivity	47
	– inhibits platelet activation and thrombosis	48
	– reduces neointima formation	49
	– overexpression protects against atherosclerosis	50
	– anti-inflammatory and anti-adhesive	51, 52
	– inhibits transformation of monocytes into macrophage foam cells	53
	– reduces SMC proliferation and migration	54
Apelin	– promotes SMC contraction and is involved in blood pressure regulation	82-84
	– prevents neointima and aneurysm formation by antagonizing angiotensin II	79
	– induces SMC proliferation	80
	– apelin or apelin receptor deficiency reduces atherosclerotic lesion formation	79, 80
	– increases endothelial NO bioavailability	79
	– stimulates hypoxia-induced EC proliferation and promotes angiogenesis	85
CRP	– promotes EC proliferation, migration and angiogenesis	17
	– promotes arterial thrombosis	18
	– accelerates the progression of atherosclerosis, but lack of proatherogenic effects has also been reported	19, 21
	– reduced neointima formation was observed in CRP transgenic mice	20
Leptin	– promotes platelet activation and thrombosis	41, 42
	– enhances neointima formation	37
	– promotes atherosclerosis	38
	– increases the angiogenic properties of EPC	33, 43
	– stimulates SMC proliferation and migration	40
MCP-1	– may mediate adipose tissue inflammation and insulin resistance in obesity	10
PAI-1	– inhibits plasminogen activation and fibrinolysis	60
	– promotes thrombus formation, neointima formation and atherosclerosis in mouse models	63, 64
	– modulates angiogenesis and cell migration	61
	– involved in the development of obesity and insulin resistance	65-71
Resistin	– interferes with the insulin-mediated cellular glucose uptake and is involved in the development of insulin resistance in obesity	72
	– inhibits adipocyte differentiation	73

CRP – C-reactive protein; EC – endothelial cells; EPC – endothelial progenitor cells; MCP-1 – monocyte chemoattractant protein-1; PAI-1 – plasminogen activator inhibitor-1; SMC – smooth muscle cells; TNF α – tumor necrosis factor- α .

factors,²⁶ suggesting that proinflammatory or vasoactive mediators from perivascular adipose depots may affect cardiovascular disease in a paracrine manner. In this regard, human aortic atherosclerosis was found to correlate with leptin, visfatin and chemerin expression in the peri-aortic adipose tissue,²⁷ and elevated perivascular expression of proinflammatory cytokines has also been reported in hypercholesterolemic or diet-induced obese mice.^{28,29} However, a direct, causal link between perivascular fat accumulation and vascular wall dysfunction is still lacking. Clearly, further studies are needed to clarify the exact role of perivascular adipose tissue and the paracrine action of adipokines or other mediators in the pathogenesis of atherosclerosis.

Vascular actions of specific adipokines

This section will briefly highlight important findings concerning the role of typical (e.g. leptin, adiponectin) and novel (e.g. apelin) adipokines on vascular cells and their effects in mouse models of atherosclerosis or intimal hyperplasia. Please also see Table 1.

Leptin

Under physiological conditions, a complex network of neuronal messenger proteins keeps our body weight relatively constant over time, within the range of a few kilograms. The adipocyte-derived hormone leptin plays an important role in this regulatory network by

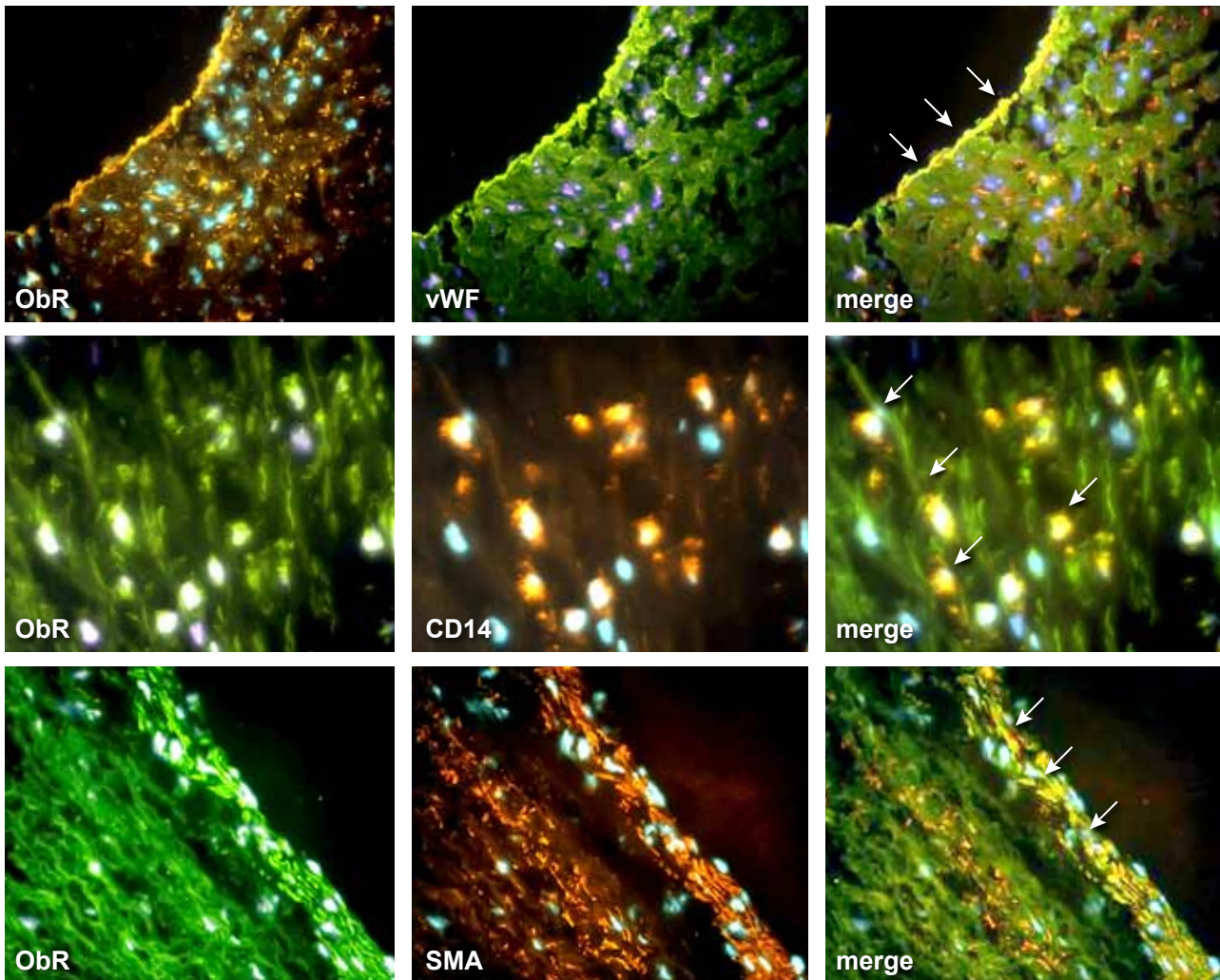


Figure 3. Expression of the leptin receptor in human atherosclerotic lesions. Double immunostaining of frozen sections through human atherosclerotic lesions for the leptin receptor (ObR) and von Willebrand factor (vWF), the LPS receptor (CD14) or smooth muscle cell α -actin (SMA), respectively, revealed its expression on endothelial cells (upper row), monocytes/macrophages (middle row) and vascular smooth muscle cells (lower row). Arrows point to double-positive cells. Magnification, 400 \times (upper and lower rows) and 630 \times (middle row).

functioning as mediator between the adipose tissue and hypothalamic neurons, controlling food intake, appetite and energy consumption.³⁰ The increase in adipose tissue mass in obesity is associated with increased circulating leptin levels;³¹ however, obese subjects are unable to respond to the weight-reducing actions of the hormone. The exact molecular mechanisms of this so-called 'leptin resistance' are not fully understood, but failure of the adipokine to cross the blood-brain barrier, impairment of leptin-mediated signal transduction on the receptor and post-receptor level, or reduction of the free, biologically active form of leptin due to binding to plasma proteins, including CRP, are being discussed.³² Importantly, weight loss may restore the cellular response to leptin, as we

were recently able to show for circulating angiogenic cells (also called early outgrowth endothelial progenitor cells), isolated from the blood of formerly obese individuals.³³ Notably, clinical studies suggested that elevated leptin levels may constitute an independent cardiovascular risk factor,³⁴ although prospective clinical trials on the importance of leptin as a mediator of the elevated cardiovascular risk in obesity have found only moderate associations.³⁵

Interestingly, the leptin receptor is expressed on a variety of cell types outside the central nervous system; these include cells present within the normal vascular wall (endothelial and smooth muscle cells), and also monocytes and T lymphocytes involved in the pathogenesis of atherosclerosis (Figure 3).³⁶ In

mice, leptin has been shown to promote neointima formation in response to vascular injury³⁷ or to enhance atherosclerotic lesion formation.³⁸ The many effects of leptin within the cardiovascular system have been reviewed in detail elsewhere,³⁹ and include enhanced proliferation and migration of smooth muscle cells⁴⁰ and the potentiation of platelet aggregation and thrombosis.^{41,42}

On the other hand, leptin may also exert potentially beneficial effects. Importantly, the hormone was found to promote the angiogenic properties of endothelial (progenitor) cells^{33,43} and to enhance the re-endothelialization of vascular lesions.⁴⁴ Thus, it remains to be shown whether the elevated cardiovascular risk in obesity is the consequence of elevated leptin levels or rather the result of resistance to leptin's potentially beneficial effects (Figure 4). In any case, the finding that an adipocyte-derived hormone, in addition to its primary mode of action (i.e. the central regulation of food intake) also exerts pleiotropic effects on vascular cells, has stimulated further research into the adipose tissue as an endocrine/paracrine organ and the vascular effects of adipokines.

Adiponectin

Adiponectin is exclusively produced in adipocytes and regulates energy metabolism and insulin sensitivity. For example, mice deficient in adiponectin exhibit severe diabetic features,⁴⁵ whereas adiponectin over-

expression⁴⁵ or replenishment⁴⁶ was found to ameliorate insulin resistance. Adiponectin receptors are expressed (among other organs) in the liver (adipoR2), skeletal muscle and heart (adipoR1), and binding of the hormone increases insulin sensitivity through inhibition of hepatic gluconeogenesis and stimulation of glucose uptake and fatty acid oxidation in muscle.⁴⁷ Thus, the reduction in circulating adiponectin levels in overweight individuals may contribute to the insulin resistance associated with obesity.

Apart from its metabolic and insulin-sensitizing activities, adiponectin also possesses direct vasculoprotective and anti-atherosclerotic properties. Studies in animal models have shown that a lack of adiponectin enhanced thrombus formation⁴⁸ and intimal hyperplasia,⁴⁹ whereas adiponectin overexpression was found to protect against atherosclerosis progression.⁵⁰ The latter observation might be related to the finding that adiponectin inhibits activation of the proinflammatory transcription factor NF κ B⁵¹ and reduces the adhesion of monocytes on activated endothelial cells,⁵² a crucial step in the initiation of vascular lesion development. Furthermore, adiponectin inhibits the transformation of monocytes into macrophage-foam cells,⁵³ and reduces the growth factor-induced proliferation and migration of vascular smooth muscle cells.⁵⁴ In accordance with these findings, persons with increased cardiovascular risk,⁵⁵ angiographically confirmed coronary heart disease,⁵⁶ or acute coronary syndromes⁵⁷ were reported to have reduced circulating adiponectin levels. However, the potential usefulness of adiponectin as biomarker for cardiovascular disease is unclear so far, possibly because determination of plasma adiponectin levels is unreliable due to the presence of multiple isoforms (at least 3 different isoforms are known to date), multimerization or binding of adiponectin to other plasma proteins.

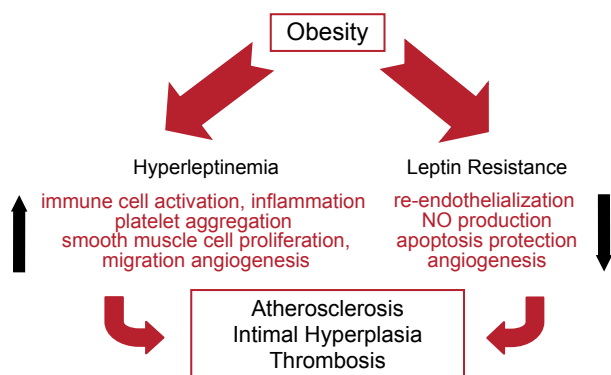


Figure 4. Hyperleptinemia and cardiovascular risk. It remains to be determined whether the increased cardiovascular risk associated with obesity occurs as consequence of a undisturbed response of vascular (e.g. endothelial and smooth muscle cells, macrophages) and hematological cells (i.e. platelets, monocytes, T-lymphocytes) to the increased circulating levels of the adipokine, and/or whether atherosclerosis initiation and progression are enhanced due to resistance to the beneficial effects of leptin.

Plasminogen activator inhibitor type-1

Plasminogen activator inhibitor type-1 (PAI-1) is produced in the adipose tissue (primarily by cells residing within the stromal cell fraction of visceral fat).⁵⁸ Elevated plasma levels of PAI-1 are frequently observed in overweight and obese subjects and decrease after weight loss.⁵⁹ PAI-1 is the principal inhibitor of urokinase and tissue-type plasminogen activator and thus a key regulator of fibrinolysis,⁶⁰ but also of other plasmin-mediated processes including activation of matrix-metalloproteinases, cell migration and angiogenesis, i.e. processes which may be of potential relevance during the evolution of vascular lesions.⁶¹

Clinical studies have revealed an association between elevated plasma PAI-1 levels and the incidence of thrombosis and atherosclerosis,⁶² while experimental studies in mice confirmed the importance of PAI-1 in the pathogenesis of the vascular complications in obesity.^{63,64} Experimental studies have also suggested a direct, causal role of PAI-1 in obesity and insulin resistance. For example, PAI-1 deficiency was found to reduce adiposity and improve the metabolic profile in both genetically and diet-induced obese mice,⁶⁵ while bone marrow transplantation studies revealed that PAI-1 expressed by macrophages within the visceral adipose tissue contributes to the development of obesity.⁶⁶ Also, pharmacological inhibition of PAI-1 dose-dependently ameliorated the development of obesity in mice that were fed a high-fat diet,⁶⁷ and prevented the development of obesity in mice,⁶⁸ whereas injection of pre-adipocytes overexpressing murine PAI-1 increased the size of *de novo* fat pads in response to a high-fat diet.⁶⁹ It should be noted that opposite findings also have been reported, namely that transgenic overexpression of PAI-1 in fat ameliorates the development of obesity in mice,⁷⁰ and that PAI-1 deficiency reduces weight gain in response to a high-fat diet.⁷¹

Resistin

Resistin (for “resistance to insulin”) was described for the first time in 2001 and was found to circulate at elevated concentrations in the plasma of obese and diabetic mice.⁷² Injection of resistin into mice produces insulin resistance, whereas administration of neutralizing antibodies improves the insulin sensitivity of obese animals.⁷³ Similarly, adipocytes in culture respond to resistin with a reduction of insulin-mediated glucose uptake.⁷² Thus, resistin could represent an important link between obesity and insulin resistance or diabetes mellitus. However, not all clinical studies were able to demonstrate elevated resistin levels in overweight persons.⁷⁴ Moreover, and in contrast to mice, human resistin is produced primarily in inflammatory cells, such as macrophages, but not in adipocytes.⁷⁵ For these reasons, findings in mice cannot be directly extrapolated to the human situation and the role of resistin in the increased cardiovascular risk associated with obesity is still not clear.

Apelin

In 1998, a small peptide was isolated from bovine stomach extracts and identified as endogenous li-

gand for the orphan G-protein-coupled receptor APJ.⁷⁶ Further studies revealed that apelin is widely expressed, including within the adipose tissue. Interestingly, its expression in adipocytes was found to increase in response to insulin or TNF α , and elevated apelin plasma levels have been described in obesity.⁷⁷ However, weight loss did not alter plasma apelin levels in obese individuals, in contrast to those of leptin and adiponectin, suggesting a weak correlation between fat mass and apelin secretion.⁷⁸ Interestingly, the receptor for apelin possesses close homologies with the angiotensin II receptor, and apelin was shown to inhibit native atherosclerosis or to prevent aneurysm and neointima formation accelerated by angiotensin II.⁷⁹ Similarly, atherosclerosis-prone mice lacking apelin receptors exhibited smaller lesions.⁸⁰ Apelin receptors are expressed on the main cellular components of the normal vascular wall,⁸¹ and apelin was shown to promote the proliferation⁸⁰ and contraction of smooth muscle cells⁸² or to regulate blood pressure.^{83,84} Also, apelin was found to promote endothelial nitric monoxide production, while hypoxia-induced apelin expression stimulated endothelial cell proliferation and angiogenesis.⁸⁵ Thus, the true or net effect of apelin on the cardiovascular system, including those in the context of increased body fat, is not fully understood at the moment.

Other adipokines

Finally, recently described adipokines include visfatin (for “visceral adipose tissue-derived serpin”), omentin and chemerin; however, their effects on vascular cells and the arterial wall in obesity are largely unknown, leaving space for future exciting discoveries in the rapidly evolving field of clinical and experimental obesity research.

Summary and future perspectives

The high prevalence of obesity, an important cardiovascular risk factor, underlines the necessity to better understand the causal relationship and interactions between the adipose tissue and the vascular wall. Adipokines, their receptors on vascular cells, and downstream intracellular signal transduction events may constitute important therapeutic target structures to modulate the effects of adipokines within the cardiovascular system, in addition to the modulation of adipose tissue inflammation or neovascularization. Along with the growing knowledge about the cellu-

lar and molecular mediators of obesity and its consequences within the cardiovascular system, our efforts should focus on the primary prevention of obesity, especially in the young, as well as the implementation of lifestyle modification strategies aiming at weight loss and increased physical activity.

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References

- Kapantais E, Tzotzas T, Ioannidis I, et al. First national epidemiological survey on the prevalence of obesity and abdominal fat distribution in Greek adults. *Ann Nutr Metab.* 2006; 50: 330-338.
- Athyros VG, Bouloukos VI, Pehlivanidis AN, et al. The prevalence of the metabolic syndrome in Greece: the MetS-Greece Multicentre Study. *Diabetes Obes Metab.* 2005; 7: 397-405.
- Haftenberger M, Lahmann PH, Panico S, et al. Overweight, obesity and fat distribution in 50- to 64-year-old participants in the European Prospective Investigation into Cancer and Nutrition (EPIC). *Public Health Nutr.* 2002; 5: 1147-1162.
- Panagiotakos DB, Pitsavos C, Chrysohoou C, et al. Epidemiology of overweight and obesity in a Greek adult population: the ATTICA Study. *Obes Res.* 2004; 12: 1914-1920.
- Eckel RH, Kahn R, Robertson RM, Rizza RA. Preventing cardiovascular disease and diabetes: a call to action from the American Diabetes Association and the American Heart Association. *Circulation.* 2006; 113: 2943-2946.
- Guh DP, Zhang W, Bansback N, Amarsi Z, Birmingham CL, Anis AH. The incidence of co-morbidities related to obesity and overweight: a systematic review and meta-analysis. *BMC Public Health.* 2009; 9: 88.
- Kenchaiah S, Evans JC, Levy D, et al. Obesity and the risk of heart failure. *N Engl J Med.* 2002; 347: 305-313.
- Stewart ST, Cutler DM, Rosen AB. Forecasting the effects of obesity and smoking on U.S. life expectancy. *N Engl J Med.* 2009; 361: 2252-2260.
- Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW. Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest.* 2003; 112: 1796-1808.
- Weisberg SP, Hunter D, Huber R, et al. CCR2 modulates inflammatory and metabolic effects of high-fat feeding. *J Clin Invest.* 2006; 116: 115-124.
- Rupnick MA, Panigrahy D, Zhang CY, et al. Adipose tissue mass can be regulated through the vasculature. *Proc Natl Acad Sci U S A.* 2002; 99: 10730-10735.
- Visser M, Bouter LM, McQuillan GM, Wener MH, Harris TB. Elevated C-reactive protein levels in overweight and obese adults. *JAMA.* 1999; 282: 2131-2135.
- Calabro P, Chang DW, Willerson JT, Yeh ET. Release of C-reactive protein in response to inflammatory cytokines by human adipocytes: linking obesity to vascular inflammation. *J Am Coll Cardiol.* 2005; 46: 1112-1113.
- Lemieux I, Pascot A, Prud'homme D, et al. Elevated C-reactive protein: another component of the atherothrombotic profile of abdominal obesity. *Arterioscler Thromb Vasc Biol.* 2001; 21: 961-967.
- Heilbronn LK, Noakes M, Clifton PM. Energy restriction and weight loss on very-low-fat diets reduce C-reactive protein concentrations in obese, healthy women. *Arterioscler Thromb Vasc Biol.* 2001; 21: 968-970.
- Ridker PM, Wilson PW, Grundy SM. Should C-reactive protein be added to metabolic syndrome and to assessment of global cardiovascular risk? *Circulation.* 2004; 109: 2818-2825.
- Turu MM, Slevin M, Matou S, et al. C-reactive protein exerts angiogenic effects on vascular endothelial cells and modulates associated signalling pathways and gene expression. *BMC Cell Biol.* 2008; 9: 47.
- Danenberg HD, Szalai AJ, Swaminathan RV, et al. Increased thrombosis after arterial injury in human C-reactive protein-transgenic mice. *Circulation.* 2003; 108: 512-515.
- Paul A, Ko KW, Li L, et al. C-reactive protein accelerates the progression of atherosclerosis in apolipoprotein E-deficient mice. *Circulation.* 2004; 109: 647-655.
- Danenberg HD, Grad E, Swaminathan RV, et al. Neointimal formation is reduced after arterial injury in human crp transgenic mice. *Atherosclerosis.* 2008; 201: 85-91.
- Hirschfield GM, Gallimore JR, Kahan MC, et al. Transgenic human C-reactive protein is not proatherogenic in apolipoprotein E-deficient mice. *Proc Natl Acad Sci U S A.* 2005; 102: 8309-8314.
- Björntorp P. “Portal” adipose tissue as a generator of risk factors for cardiovascular disease and diabetes. *Arteriosclerosis.* 1990; 10: 493-496.
- Fain JN, Madan AK, Hiler ML, Cheema P, Bahouth SW. Comparison of the release of adipokines by adipose tissue, adipose tissue matrix, and adipocytes from visceral and subcutaneous abdominal adipose tissues of obese humans. *Endocrinology.* 2004; 145: 2273-2282.
- Rosito GA, Massaro JM, Hoffmann U, et al. Pericardial fat, visceral abdominal fat, cardiovascular disease risk factors, and vascular calcification in a community-based sample: the Framingham Heart Study. *Circulation.* 2008; 117: 605-613.
- Mahabadi AA, Massaro JM, Rosito GA, et al. Association of pericardial fat, intrathoracic fat, and visceral abdominal fat with cardiovascular disease burden: the Framingham Heart Study. *Eur Heart J.* 2009; 30: 850-856.
- Mahabadi AA, Reinsch N, Lehmann N, et al. Association of pericoronary fat volume with atherosclerotic plaque burden in the underlying coronary artery: A segment analysis. *Atherosclerosis.* 2010; 211: 195-199.
- Spiroglou SG, Kostopoulos CG, Varakis JN, Papadaki HH. Adipokines in periaortic and epicardial adipose tissue: differential expression and relation to atherosclerosis. *J Atheroscler Thromb.* 2010; 17: 115-130.
- Lohmann C, Schäfer N, von Lukowicz T, et al. Atherosclerotic mice exhibit systemic inflammation in periaortic and visceral adipose tissue, liver, and pancreatic islets. *Atherosclerosis.* 2009; 207: 360-367.

29. Henrichot E, Juge-Aubry CE, Pernin A, et al. Production of chemokines by perivascular adipose tissue: a role in the pathogenesis of atherosclerosis? *Arterioscler Thromb Vasc Biol.* 2005; 25: 2594-2599.
30. Friedman JM, Halaas JL. Leptin and the regulation of body weight in mammals. *Nature.* 1998; 395: 763-770.
31. Maffei M, Halaas J, Ravussin E, et al. Leptin levels in human and rodent: measurement of plasma leptin and ob RNA in obese and weight-reduced subjects. *Nat Med.* 1995; 1: 1155-1161.
32. Münzberg H, Myers MG. Molecular and anatomical determinants of central leptin resistance. *Nat Neurosci.* 2005; 8: 566-570.
33. Heida NM, Leifheit-Nestler M, Schroeter MR, et al. Leptin enhances the potency of circulating angiogenic cells via src kinase and integrin (alpha) vbeta 5: implications for angiogenesis in human obesity. *Arterioscler Thromb Vasc Biol.* 2010; 30: 200-206.
34. Leyva F, Godsland IF, Ghatel M, et al. Hyperleptinemia as a component of a metabolic syndrome of cardiovascular risk. *Arterioscler Thromb Vasc Biol.* 1998; 18: 928-933.
35. Sattar N, Wannamethee G, Sarwar N, et al. Leptin and coronary heart disease: prospective study and systematic review. *J Am Coll Cardiol.* 2009; 53: 167-175.
36. Schroeter MR, Schneiderman J, Schumann B, et al. Expression of the leptin receptor in different types of vascular lesions. *Histochem Cell Biol.* 2007; 128: 323-333.
37. Schäfer K, Halle M, Goeschen C, et al. Leptin promotes vascular remodeling and neointimal growth in mice. *Arterioscler Thromb Vasc Biol.* 2004; 24: 112-117.
38. Bodary PF, Gu S, Shen Y, Hasty AH, Buckler JM, Eitzman DT. Recombinant leptin promotes atherosclerosis and thrombosis in apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol.* 2005; 25: e119-122.
39. Beltowski J. Leptin and atherosclerosis. *Atherosclerosis.* 2006; 189: 47-60.
40. Oda A, Taniguchi T, Yokoyama M. Leptin stimulates rat aortic smooth muscle cell proliferation and migration. *Kobe J Med Sci.* 2001; 47: 141-150.
41. Konstantinides S, Schäfer K, Koschnick S, Loskutoff DJ. Leptin-dependent platelet aggregation and arterial thrombosis suggests a mechanism for atherothrombotic disease in obesity. *J Clin Invest.* 2001; 108: 1533-1540.
42. Bodary PF, Westrick RJ, Wickenheiser KJ, Shen YC, Eitzman DT. Effect of leptin on arterial thrombosis following vascular injury in mice. *JAMA.* 2002; 287: 1706-1709.
43. Sierra-Honigmann MR, Nath AK, Murakami C, et al. Biological action of leptin as an angiogenic factor. *Science.* 1998; 281: 1683-1686.
44. Schroeter MR, Leifheit M, Sudholt P, et al. Leptin enhances the recruitment of endothelial progenitor cells into neointimal lesions after vascular injury by promoting integrin-mediated adhesion. *Circ Res.* 2008; 103: 536-544.
45. Maeda N, Shimomura I, Kishida K, et al. Diet-induced insulin resistance in mice lacking adiponectin/ACRP30. *Nat Med.* 2002; 8: 731-737.
46. Yamauchi T, Kamon J, Waki H, et al. The fat-derived hormone adiponectin reverses insulin resistance associated with both lipodystrophy and obesity. *Nat Med.* 2001; 7: 941-946.
47. Yamauchi T, Kamon J, Minokoshi Y, et al. Adiponectin stimulates glucose utilization and fatty-acid oxidation by activating AMP-activated protein kinase. *Nat Med.* 2002; 8: 1288-1295.
48. Kato H, Kashiwagi H, Shiraga M, et al. Adiponectin acts as an endogenous antithrombotic factor. *Arterioscler Thromb Vasc Biol.* 2006; 26: 224-230.
49. Kubota N, Terauchi Y, Yamauchi T, et al. Disruption of adiponectin causes insulin resistance and neointimal formation. *J Biol Chem.* 2002; 277: 25863-25866.
50. Okamoto Y, Kihara S, Ouchi N, et al. Adiponectin reduces atherosclerosis in apolipoprotein E-deficient mice. *Circulation.* 2002; 106: 2767-2770.
51. Ouchi N, Kihara S, Arita Y, et al. Adiponectin, an adipocyte-derived plasma protein, inhibits endothelial NF-kappaB signaling through a cAMP-dependent pathway. *Circulation.* 2000; 102: 1296-1301.
52. Ouchi N, Kihara S, Arita Y, et al. Novel modulator for endothelial adhesion molecules: adipocyte-derived plasma protein adiponectin. *Circulation.* 1999; 100: 2473-2476.
53. Ouchi N, Kihara S, Arita Y, et al. Adipocyte-derived plasma protein, adiponectin, suppresses lipid accumulation and class A scavenger receptor expression in human monocyte-derived macrophages. *Circulation.* 2001; 103: 1057-1063.
54. Arita Y, Kihara S, Ouchi N, et al. Adipocyte-derived plasma protein adiponectin acts as a platelet-derived growth factor-BB-binding protein and regulates growth factor-induced common postreceptor signal in vascular smooth muscle cell. *Circulation.* 2002; 105: 2893-2898.
55. Pischon T, Girman CJ, Hotamisligil GS, Rifai N, Hu FB, Rimm EB. Plasma adiponectin levels and risk of myocardial infarction in men. *JAMA.* 2004; 291: 1730-1737.
56. Ouchi N, Kihara S, Funahashi T, et al. Reciprocal association of C-reactive protein with adiponectin in blood stream and adipose tissue. *Circulation.* 2003; 107: 671-674.
57. Wolk R, Berger P, Lennon RJ, Brilakis ES, Davison DE, Somers VK. Association between plasma adiponectin levels and unstable coronary syndromes. *Eur Heart J.* 2007; 28: 292-298.
58. Bastelica D, Morange P, Berthet B, et al. Stromal cells are the main plasminogen activator inhibitor-1-producing cells in human fat: evidence of differences between visceral and subcutaneous deposits. *Arterioscler Thromb Vasc Biol.* 2002; 22: 173-178.
59. Kockx M, Leenen R, Seidell J, Princen HM, Kooistra T. Relationship between visceral fat and PAI-1 in overweight men and women before and after weight loss. *Thromb Haemost.* 1999; 82: 1490-1496.
60. Loskutoff DJ, Samad F. The adipocyte and hemostatic balance in obesity: studies of PAI-1. *Arterioscler Thromb Vasc Biol.* 1998; 18: 1-6.
61. Binder BR, Mihaly J, Prager GW. uPAR-uPA-PAI-1 interactions and signaling: a vascular biologist's view. *Thromb Haemost.* 2007; 97: 336-342.
62. Juhan-Vague I, Alessi MC. PAI-1, obesity, insulin resistance and risk of cardiovascular events. *Thromb Haemost.* 1997; 78: 656-660.
63. Schäfer K, Müller K, Hecke A, et al. Enhanced thrombosis in atherosclerosis-prone mice is associated with increased arterial expression of plasminogen activator inhibitor-1. *Arterioscler Thromb Vasc Biol.* 2003; 23: 2097-2103.
64. Konstantinides S, Schäfer K, Thinnis T, Loskutoff DJ. Plasminogen activator inhibitor-1 and its cofactor vitronectin stabilize arterial thrombi after vascular injury in mice. *Circulation.* 2001; 103: 576-583.
65. Schäfer K, Fujisawa K, Konstantinides S, Loskutoff DJ. Disruption of the plasminogen activator inhibitor 1 gene reduces the adiposity and improves the metabolic profile of genetic-

- ly obese and diabetic ob/ob mice. *FASEB J*. 2001; 15: 1840-1842.
66. De Tacey BM, Novitskaya T, Gleaves L, Covington JW, Vaughan DE. Bone marrow plasminogen activator inhibitor-1 influences the development of obesity. *J Biol Chem*. 2006; 281: 32796-32805.
 67. Ma LJ, Mao SL, Taylor KL, et al. Prevention of obesity and insulin resistance in mice lacking plasminogen activator inhibitor 1. *Diabetes*. 2004; 53: 336-346.
 68. Crandall DL, Quinet EM, El Ayachi S, et al. Modulation of adipose tissue development by pharmacological inhibition of PAI-1. *Arterioscler Thromb Vasc Biol*. 2006; 26: 2209-2215.
 69. Scroyen I, Jacobs F, Cosemans L, De Geest B, Lijnen HR. Effect of plasminogen activator inhibitor-1 on adipogenesis in vivo. *Thromb Haemost*. 2009; 101: 388-393.
 70. Lijnen HR, Maquoi E, Morange P, et al. Nutritionally induced obesity is attenuated in transgenic mice overexpressing plasminogen activator inhibitor-1. *Arterioscler Thromb Vasc Biol*. 2003; 23: 78-84.
 71. Morange PE, Lijnen HR, Alessi MC, Kopp F, Collen D, Juhan-Vague I. Influence of PAI-1 on adipose tissue growth and metabolic parameters in a murine model of diet-induced obesity. *Arterioscler Thromb Vasc Biol*. 2000; 20: 1150-1154.
 72. Stepan CM, Bailey ST, Bhat S, et al. The hormone resistin links obesity to diabetes. *Nature*. 2001; 409: 307-312.
 73. Kim KH, Lee K, Moon YS, Sul HS. A cysteine-rich adipose tissue-specific secretory factor inhibits adipocyte differentiation. *J Biol Chem*. 2001; 276: 11252-11256.
 74. Lee JH, Chan JL, Yiannakouris N, et al. Circulating resistin levels are not associated with obesity or insulin resistance in humans and are not regulated by fasting or leptin administration: cross-sectional and interventional studies in normal, insulin-resistant, and diabetic subjects. *J Clin Endocrinol Metab*. 2003; 88: 4848-4856.
 75. Jung HS, Park KH, Cho YM, et al. Resistin is secreted from macrophages in atheromas and promotes atherosclerosis. *Cardiovasc Res*. 2006; 69: 76-85.
 76. Tatemoto K, Hosoya M, Habata Y, et al. Isolation and characterization of a novel endogenous peptide ligand for the human APJ receptor. *Biochem Biophys Res Commun*. 1998; 251: 471-476.
 77. Boucher J, Masri B, Daviaud D, et al. Apelin, a newly identified adipokine up-regulated by insulin and obesity. *Endocrinology*. 2005; 146: 1764-1771.
 78. Heinonen MV, Laaksonen DE, Karhu T, et al. Effect of diet-induced weight loss on plasma apelin and cytokine levels in individuals with the metabolic syndrome. *Nutr Metab Cardiovasc Dis*. 2009; 19: 626-633.
 79. Chun HJ, Ali ZA, Kojima Y, et al. Apelin signaling antagonizes Ang II effects in mouse models of atherosclerosis. *J Clin Invest*. 2008; 118: 3343-3354.
 80. Hashimoto T, Kihara M, Imai N, et al. Requirement of apelin-apelin receptor system for oxidative stress-linked atherosclerosis. *Am J Pathol*. 2007; 171: 1705-1712.
 81. Kleinz MJ, Skepper JN, Davenport AP. Immunocytochemical localisation of the apelin receptor, APJ, to human cardiomyocytes, vascular smooth muscle and endothelial cells. *Regul Pept*. 2005; 126: 233-240.
 82. Hashimoto T, Kihara M, Ishida J, et al. Apelin stimulates myosin light chain phosphorylation in vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol*. 2006; 26: 1267-1272.
 83. Zhang Q, Yao F, Raizada MK, O'Rourke ST, Sun C. Apelin gene transfer into the rostral ventrolateral medulla induces chronic blood pressure elevation in normotensive rats. *Circ Res*. 2009; 104: 1421-1428.
 84. Ishida J, Hashimoto T, Hashimoto Y, et al. Regulatory roles for APJ, a seven-transmembrane receptor related to angiotensin-type 1 receptor in blood pressure in vivo. *J Biol Chem*. 2004; 279: 26274-26279.
 85. Eyries M, Siegfried G, Ciumas M, et al. Hypoxia-induced apelin expression regulates endothelial cell proliferation and regenerative angiogenesis. *Circ Res*. 2008; 103: 432-440.